

Current findings on terrestrial plants – Engineered nanomaterial interactions: Are plants capable of phytoremediating nanomaterials from soil?

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Abstract

Engineered Nanomaterials (ENMs) are revolutionizing our daily lives, industry, and agriculture. Along with their novel applications, major concerns have emerged due to the potential toxicity to biological systems. Since soils are considered the main destination for ENMs, research focused on their interaction with plants is gaining more attention, especially at the physiological and biochemical levels. This review addresses the capacity of some plants to accumulate ENMs or released ions, highlighting the beneficial and detrimental effects and the potential use of some plants to remediate ENM-contaminated environments. Although the uptake process depends on multiple factors, the literature suggests that concentrations <50 mg/kg are beneficial, while higher doses negatively impact physiological and biochemical parameters. However, the current data does not allow the formulation of mechanistic model effects. Finally, this review remarks on the pivotal role played by plants as a sustainable alternative to face the environmental buildup of ENMs and to guarantee food security.

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Introduction

Nanotechnology is an emerging field where scientific knowledge is used to manipulate matter at the nano-scale (10^{-9} m) [1]. The nanotechnology industry has grown exponentially through the use of Engineered Nanomaterials (ENMs). ENMs have vast applications in superconductors, next generation medicine, automotive, electronics, sporting goods, environmental remediation, food, and agriculture, among others [2–5] (Figure 1). ENMs include carbon-based, metal-based, quantum dots, and others [6,7]. Notably, pros and cons of the ENM production/use are estimated to go beyond those caused by the industrial revolution [8].

Soils are the main repository for ENMs, which directly impact terrestrial plants [9] (Figure 1). ENMs are incorporated in agricultural soils directly through agricultural intended products, or indirectly through biosolids [10]. Estimates indicate that the concentration of titanium dioxide (TiO_2) ENM in sewage sludge ranges between 107 and 802 mg/kg [11], which could end in agricultural soils. This suggests that, in the near future, mechanisms for removing ENMs from soil may be needed. Phytoremediation, the removal of contaminants using plants, could be an alternative to reducing the environmental buildup of ENMs. This technique involves plants with/without symbiont microorganisms and includes the extraction, immobilization, or degradation of contaminants [12].

Due to their small size and high surface reactivity, ENMs are able to go inside plant cells, eliciting detrimental or positive effects [7]. This review highlights contrasting effects of ENMs in plants, which are determined by multiple factors such as plant species, microbial soil community, soil properties, and nutrient availability, among others. The ability of some plants to accumulate or degrade nano-based materials, which could be potentially used in phytoremediation, are discussed.

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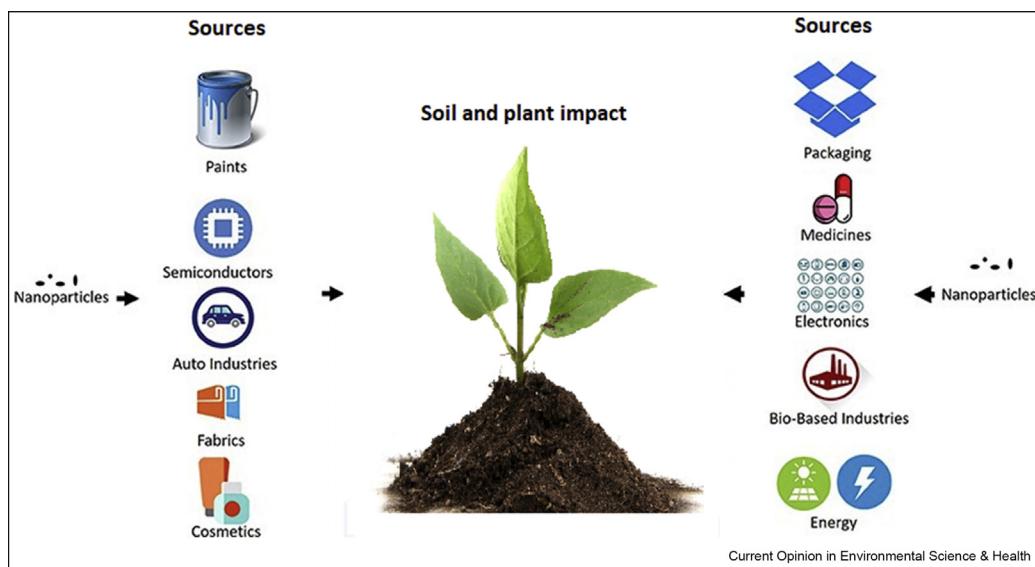
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Figure 1



Different uses of ENMs and potential impact in soil and plants after end user application. Taken and modified from Ref. [17].

A brief summary of ENMs

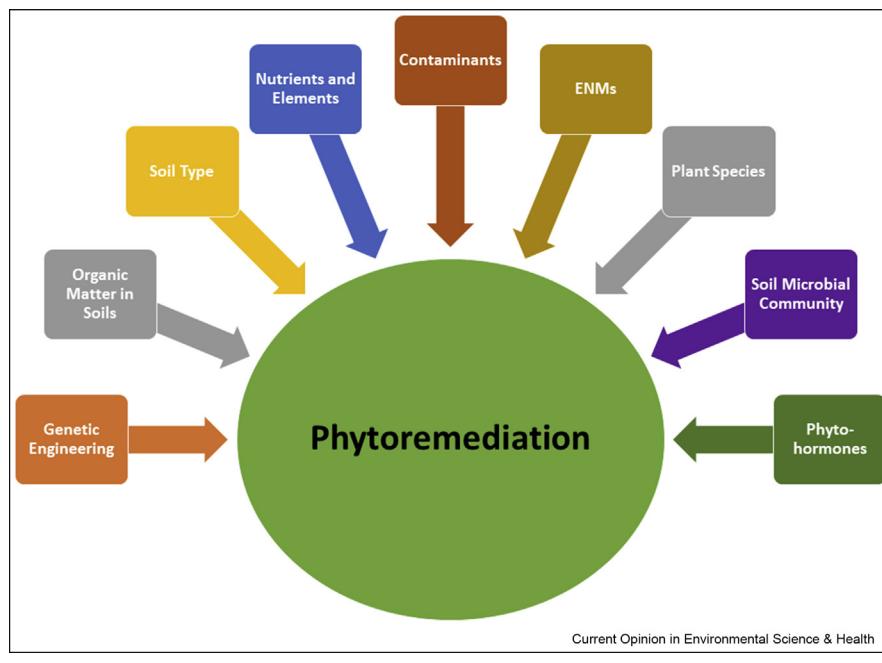
Historically, plants have been exposed to low concentrations of natural nanoparticles [13]. The industrial production and use of ENMs have changed the panorama. In the near future, plants will be exposed to carbon-based nanostructures (e.g. fullerenes, SWCNT, and MWCNT) and metal-based nanostructures [14–17]. Carbon-based nanoparticles (NPs) are used in agriculture, biomedicine, optics, and electronics [15], while metal-based ENMs are widely used as catalysts (Fe nanoparticles) [13], as antimicrobial agents (Ag NPs) [18], in agriculture (Zn and Cu-based NPs) [19], and as UV blockers (TiO_2 and ZnO NPs) [20]. Zero-valent iron NPs are gaining attention in the field of environmental remediation [21]. However, concerns regarding their potential ecotoxicity have emerged [22].

ENMs and terrestrial plants: from physiological aspects to phytoremediation

Several studies have shown that ENMs affect plants either positively and negatively, but they also suggest the possibility of soil restoration using plants. Probabilistic models have been developed to evaluate the concentration of ENMs in environmental samples. Gottschalk et al. [23] summarized the literature on this topic. A modeling study in the San Francisco Bay area suggests that, within a few years, the concentration of ENMs will pose a threat to the organisms inhabiting the Bay [20]. Thus, more research focused on understanding the relationships between ENMs and biotic/abiotic systems, especially those related to the phytoremediation potential, are needed [24,25]. This section discusses the latest findings on plant system-ENM interactions and the removal of nanostructures from soils.

Some plants possess the ability to store heavy metals at high concentrations; these plants are called hyperaccumulators [26]. Hyperaccumulation/degradation of contaminants is one of the basis of phytoremediation [27] and subsequent recovery of soil productivity [28]. Figure 2 shows that many factors may affect the phytoremediation of ENMs. The interaction of these multiple variables on phytoremediation performance is currently a very stimulating field of research. A deeper knowledge in this area will be contributory to control multiple aspects of the plant physiology such as NPs phytotoxicity and to manipulate the bioaccumulation of ENMs. For instance, in *Arabidopsis thaliana*, the use of TiO_2 NPs has demonstrated to reduce phytotoxic effects caused by tetracycline [29]. In Lettuce (*Lactuca sativa*) roots, Cu content from Cu-NPs can be 2.15-fold increased by weathering the NPs contaminated-soils [30]. Interestingly, the phytotoxic effects of ZnO NPs in maize (*Zea mays*) can be alleviated by the inoculation with mycorrhizal fungi and phosphorus supplemental [31]. The knowledge of phytoremediating ENMs has recently been accumulated. However, there is a need to increase our research on the interaction of “conventional” contaminants with NPs in phytoremediation systems to have better insights into what occurs in nature.

Once ENMs are spread in the environment, they experience transformations. By interacting with soil organic matter, these tiny particles can undergo aggregation, a process that reduces their reactivity and increases their stability. On the other hand, organic matter sometimes can cause an increment of ENM solubility, which potentiates ion release, most of the time with

Figure 2

Factors affecting phytoremediation effectiveness.

toxic repercussions on living systems [32]. Additionally, the hydrophobic properties of some ENMs (Fullerenes and MWCNT) confer on them the ability to interact with cell organic molecules, facilitating their possible intake into the cytoplasm [33,34]. This mechanism might represent an advantageous strategy for the phytoremediation of hydrophobic co-contaminants, which has been less studied.

Soil microorganisms (MOs) may also alter the response of plants to ENM exposure. MOs play fundamental roles in the dynamics and availability of macro and micro-nutrients [35]. To date, few and contrasting results have been published about the interaction of ENMs on MOs-plant symbiosis. Tomato plants (*Solanum lycopersicum* L.), inoculated with mycorrhizal fungi, stored 14% less silver than their non-mycorrhizal counterparts when challenged with 36 mg/kg of nAg [36]. In *Medicago truncatula*, the symbiotic relationship with the nitrogen-fixing bacterium, *Sinorhizobium meliloti*, was unaffected by nAg [37]. However, most of the studies have been performed under controlled conditions assessing one or a small group of MO's species. As a result, there is the possibility that the observed responses will vary in natural environments.

Phytoremediation of metal-based ENMs

Several plants have shown abilities to accumulate ENMs or the respective ions (Table 1). Many of the ENMs used in agriculture are metal-based nano-forms [38],

due to that, much of the following discussion is concentrated on these nanomaterials.

Zinc oxide (ZnO) and silver (Ag) NPs have been widely used to evaluate plant-ENMs interactions. For example, soybean plants cultivated in farm soil amended with 500 mg ZnO NPs/kg, fixed nitrogen in a similar way that the respective controls, although with some leaf damage [9]. Alfalfa (*Medicago sativa*), exposed up to 750 mg/kg of ZnO NPs showed great capability for Zn accumulation in all tissues and nodules, and also showed less toxicity as compared to ZnCl₂ treatment [16]. Chen et al. exposed the model legume *M. truncatula* to 50 mg/kg of nZnO and 5 mg/kg of nAg in soil amended with biosolids [39]. These authors found no effects on the fresh shoot and root biomass, shoot length, and root diameter, which suggests this species tolerate nZnO and nAg ENMs at lower concentrations of 50 mg/kg. Contrarily, the physiology of *A. thaliana* was affected by >300 mg Ag NPs/L, in a gel medium [40], indicating that *A. thaliana* does not tolerate Ag NPs at above-mentioned concentrations.

Cerium oxide (CeO₂) ENMs have shown to be very stable in natural environments. Thus, plants accumulate Ce, mostly as NPs. Mesquite (*Prosopis juliflora-velutina*), a desert plant, was exposed to 4000 mg CeO₂ NPs/kg without causing apparent signs of toxicity. The Ce concentration in roots was about 3600 mg/kg dry weight (DW), most of them as NPs, as it was demonstrated by synchrotron studies [41]. This suggests that mesquite is

Table 1

Summary of studies showing the phytoremediation potential of some plants due to their ability to accumulate ENMs/released ions.^a

Plant Species	Type of ENMs	Exposed Concentration	Exposed Time	Leaves Concentration (mg/kg tissue)	Shoot/Stem Concentration (mg/kg tissue)	Root Concentration (mg/kg tissue)	Medium	Reference
<i>Ocimum basilicum</i> L.	nAg	80 mg/kg	4 weeks	1.4	2.1	5.8	Soil	[57]
	nCo	80 mg/kg		3.3	2.8	71.4		
	nNi	80 mg/kg		3.8	0.6	27.3		
<i>Origanum vulgare</i>	nCu	50 mg/kg	60 days	22	23	50	Soil	[58]
<i>Glycine max</i> L. Merr.	nCeO ₂	500 mg/kg	30 days	3.5	1100	—	Soil	[42]
<i>Lactuca sativa</i> L.	nZnO	100 mg/kg	7 weeks	23	—	—	Soil	[59]
<i>Raphanus sativus</i> L.	nCeO	10 mg/L	5 days	—	0.15	12	Hydroponics	[60]

^a The reported values correspond to the total metal concentration found in tissues.

a good candidate for the remediation of CeO₂ ENMs. Rossi et al. [42] demonstrated that soybean accumulated up to 3.5 mg Ce/kg dry leaves in plants exposed to 500 mg CeO₂ NPs/kg soil, in the presence of 1 mg Cd/kg soil. Although the study did not include the use of X-ray absorption spectroscopy, it is very likely that most of the Ce in tissues was in the form of NPs. It is worth noting that the presence of the co-contaminant increased the uptake and translocation of CeO₂ NPs, and the latter did not augment Cd toxicity in the soybean.

Very few studies have shown the concentration of TiO₂ ENMs in plant tissues, because these NPs are difficult to be digested and thus analyzed. However, TiO₂ ENMs have been detected in plants using several analytical techniques [43]. Servin et al. [44], demonstrated that TiO₂ ENMs, even at 4000 mg/L, in hydroponics, increased cucumber shoot growth. Using synchrotron studies, they demonstrated that cucumber absorbed and translocated the TiO₂ NPs. It is interesting to point out that plants were fed with a mixture of anatase/rutile; however, the aerial tissues had the TiO₂ NPs mainly in the rutile form. Larue et al. [45] reported that lettuce (*L. sativa*) hydroponically exposed to 1000 mg TiO₂ NPs/L accumulated up to 99 mg/kg in the vascular system. Similarly, an experiment with *M. truncatula* demonstrated that low doses (50 mg/kg) of TiO₂ NPs did not affect the physiological parameters or plant health [39]. More recently, Tan et al. [46] cultivated basil (*Ocimum basilicum*) in soil amended with three types of TiO₂ ENMs, at 750 mg/kg, and found about 30 mg Ti/kg DW in shoots. None of the above-cited plants showed apparent toxicity, which suggests that these plants may have potential to remove TiO₂ from the soil.

Plant responses to lower ENMs exposure

There exists a vast increment in research publications focused to evaluate the effect of different types of ENMs on cultivars. Comprehensive reviews that summarize the effects of NPs on plants have been reported

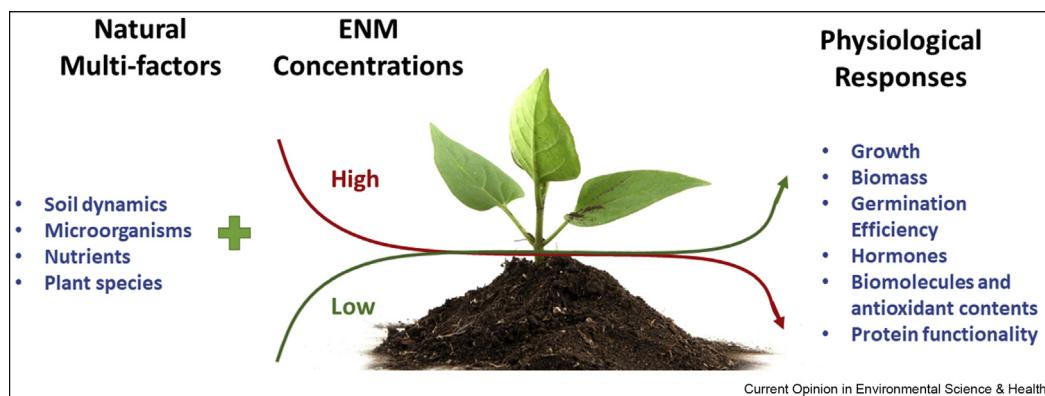
[6,17,47]. Although plant responses depend on multi-factors, some beneficial responses that have been documented, rely on ENMs type and concentration. For instance, the two weeks foliar application of ZnO NPs in the Clusterbean (*Cyamopsis tetragonoloba* L.) at 10 mg/L, augmented the young plant biomass, protein, and chlorophyll content [48]. In Parsley (*Petroselinum crispum*), the use of TiO₂ NPs at doses ranging from 10 to 40 mg/L increased fresh weight, as well as root and shoot length in seedlings [49]. In another study, the use of Fe/Fe₃O₄ NPs did not affect physiological parameters in lettuce hydroponically incubated with at 10–20 mg/L of the NPs [50]. Also, exposure of Fenugreek (*Trigonella foenum-graecum* L.) to low amounts of Ag-NPs (1 mg/L) showed to greatly improve the growth of the young plant [51]. Although it is difficult to establish a mechanistic model about the effects of ENMs on plant physiology [21,52], these examples suggest that low concentration of ENMs (<50 mg/kg or mg/L) do not affect or show positive physiological effects, while elevated concentrations inhibit plant growth [53] (Figure 3).

However, some ENMs such as CeO₂ and TiO₂ are tolerated by plants at high concentrations [41,44,46]. Additionally, the presence of coating agents (molecules covering NP's surfaces minimizing ENMs aggregation), alter the physiological responses giving contradictory results [46,54,55]. This clearly shows that the possibility of a simple answer to the exposure of plants to ENMs is difficult.

Conclusions and future perspectives

Experimental data has shown that plant species such as mesquite, alfalfa, or cucumber have the ability to uptake ENMs. Nevertheless, more studies about biochemical, physiological, molecular, and ecological levels are required to build a comprehensive model for the phytoremediation of specific pollutants. Additionally, few examples have shown that plants and plant-microorganism systems have the potential for ENMs

Figure 3



Effect of ENMs on physiological responses of plants. Albeit several factors are involved in physiological responses of plants under ENMs stress, in general, low doses (<50 mg/kg) stimulate positive physiological responses, while higher concentrations cause damages.

uptake. With the advent of genetically modified organisms (GMO), new scenarios should be taken into consideration in the discussion of food safety and environmental health. Some plants can be genetically modified to increase their ENMs hyperaccumulation [56]. In addition, genetically modified microorganisms, able to withstand high concentrations of ENMs, can be associated with plants and used in ENMs phytoremediation. However, ecological implications, such as damage to native species and interactions with other organisms should not be discarded. Metagenomic studies have to be conducted to understand the behavior, relationship, and dynamics of ecosystems. Certainly, the use of cutting-edge techniques such as next-generation sequencing (NGS) will be of paramount importance to assess soil microbial dynamics and their determinant roles for plant physiology under ENMs contaminated environments.

Conflict of interest

None.

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References

Papers of particular interest, published within the period of review, have been highlighted as:

- * of special interest
 - ** of outstanding interest
1. Bhushan B: **Introduction to nanotechnology.** In *Springer handbook of nanotechnology*. Edited by Bhushan B, Berlin Heidelberg: Springer; 2017:1–19.
 2. Giese B, Klaessig F, Park B, Kaegi R, Steinfeldt M, Wigger H, von Gleich A, Gottschalk F: **Risks, release and concentrations of engineered nanomaterial in the environment.** *Sci Rep* 2018, **8**: 1565.
 3. Pradhan S, Mailapalli DR: **Interaction of engineered nanoparticles with the agri-environment.** *J Agric Food Chem* 2017, **65**:8279–8294.
 4. Caballero-Guzman A, Nowack B: **A critical review of engineered nanomaterial release data: are current data useful for material flow modeling?** *Environ Pollut* 2016, **213**:502–517.
 5. Keller AA, McFerran S, Lazareva A, Suh S: **Global life cycle releases of engineered nanomaterials.** *J Nanoparticle Res* 2013, **15**:1692.
 6. Zuverza-Mena N, Martínez-Fernández D, Du W, Hernandez-Viecas JA, Bonilla-Bird N, López-Moreno ML, Komárek M, Peralta-Videa JR, Gardea-Torresdey JL: **Exposure of engineered nanomaterials to plants: insights into the physiological and biochemical responses-a review.** *Plant Physiol Biochem* 2017, **110**:236–264.
 7. Hatami M, Kariman K, Ghorbanpour M: **Engineered nanomaterial-mediated changes in the metabolism of terrestrial plants.** *Sci Total Environ* 2016, **571**:275–291.
 8. Navarro E, Baun A, Behra R, Hartmann NB, Filser J, Miao AJ, Quigg A, Santschi PH, Sigg L: **Environmental behavior and ecotoxicity of engineered nanoparticles to algae, plants, and fungi.** *Ecotoxicology* 2008, **17**:372–386.
 9. Priester JH, Moritz SC, Espinosa K, Ge Y, Wang Y, Nisbet RM, Schimel JP, Susana Goggi A, Gardea-Torresdey JL, Holden PA: **Damage assessment for soybean cultivated in soil with either CeO₂ or ZnO manufactured nanomaterials.** *Sci Total Environ* 2017, **579**:1756–1768.
 10. Gardea-Torresdey JL, Rico CM, White JC: **Trophic transfer, transformation, and impact of engineered nanomaterials in terrestrial environments.** *Environ Sci Technol* 2014, **48**: 2526–2540.

11. Gottschalk F, Sonderer T, Scholz RW, Nowack B: Modeled environmental concentrations of engineered nanomaterials (TiO_2 , ZnO , Ag, CNT, fullerenes) for different regions. *Environ Sci Technol* 2009, 43:9216–9222.
12. Panchenko L, Muratova A, Turkovskaya O: Comparison of the phytoremediation potentials of *Medicago falcata* L. and *Medicago sativa* L. in aged oil-sludge-contaminated soil. *Environ Sci Pollut Res* 2017, 24:3117–3130.
- This is a comprehensive study on the phytoremediation capacity of *Medicago falcata* L. and *Medicago sativa* L. Authors characterized petroleum contaminated soils and evaluated the prevalence of *Medicago* species in such ecosystem. By performing laboratory pot experiments, they demonstrated the removal of up to 18% of major heavy oil fractions. Additionally, a description of important species of microorganisms involved in hydrocarbons removal along to enzymatic assays of both soil and roots was performed.
13. Zaytseva O, Neumann G: Carbon nanomaterials: production, impact on plant development, agricultural and environmental applications. *Chem Biol Technol Agric* 2016, 3:1–26.
14. Sankar R, Manikandan P, Malarvizhi V, Fathima T, Shivashangari KS, Ravikumar V: Green synthesis of colloidal copper oxide nanoparticles using *Carica papaya* and its application in photocatalytic dye degradation. *Spectrochim Acta - Part A Mol Biomol Spectrosc* 2014, 121:746–750.
15. de la Rosa G, García-Castañeda C, Vázquez-Núñez E, Alonso-Castro ÁJ, Basurto-Islas G, Mendoza A, Cruz-Jiménez G, Molina C: Physiological and biochemical response of plants to engineered NMs: implications on future design. *Plant Physiol Biochem* 2017, 110:226–235.
- This review is focused on analyzing the physiological effects of cerium, copper, and iron nano-particles (NPs) on different plant species. Authors described different NPs exposure paths for plant systems, followed by a deep, current literature discussion on the interaction of NPs, plants and microorganisms. Since the physico-chemical properties of NPs and plant species determine the uptake of the NPs, authors concluded that this knowledge should be taken into consideration for future NP designs.
16. Bandyopadhyay S, Plascencia-Villa G, Mukherjee A, Rico CM, José-Yacamán M, Peralta-Videa JR, Gardea-Torresdey JL: Comparative phytotoxicity of ZnO NPs, bulk ZnO , and ionic zinc onto the alfalfa plants symbiotically associated with *Sinorhizobium meliloti* in soil. *Sci Total Environ* 2015, 515–516:60–69.
17. Rastogi A, Zivcak M, Sytar O, Kalaji HM, He X, Mbarki S, Brestic M: Impact of metal and metal oxide nanoparticles on plant: a critical review. *Front Chem* 2017, 5:1–16.
18. Ma X, Geisler-Lee J, Deng Y, Kolmakov A: Interactions between engineered nanoparticles (ENPs) and plants: phytotoxicity, uptake and accumulation. *Sci Total Environ* 2010, 408: 3053–3061.
19. Antonoglou O, Moustaka J, Adamakis ID, Sperdouli I, Pantazaki AA, Moustakas M, Dendrinou-Samara C: Nanobrass CuZn nanoparticles as foliar spray non phytotoxic fungicides. *ACS Appl Mater Interfaces* 2018, 10:4450–4461.
20. Garner KL, Suh S, Keller AA: Assessing the risk of engineered nanomaterials in the environment: development and application of the nanoFate model. *Environ Sci Technol* 2017, 51: 5541–5551.
21. Montes A, Bisson MA, Gardella JA, Aga DS: Uptake and transformations of engineered nanomaterials: critical responses observed in terrestrial plants and the model plant *Arabidopsis thaliana*. *Sci Total Environ* 2017, 607–608:1497–1516.
- This article documents the interaction of plants and engineered nanomaterials (ENMs) by highlighting the importance of using inductively coupled plasma mass spectrometry (ICP-MS) and imaging-based techniques to investigate ENMs/ions uptake, translocation, and accumulation in plants. Very good discussion on the physiological, biochemical, and genetic effects of the type, size, and ENM concentration in *Arabidopsis thaliana* is presented. Authors recommend the use of this model plant to gain deeper knowledge about the interactions of ENMs and plants.
22. Ma X, Gurung A, Deng Y: Phytotoxicity and uptake of nano-scale zero-valent iron (nZVI) by two plant species. *Sci Total Environ* 2013, 443:844–849.
23. Gottschalk F, Sun T, Nowack B: Environmental concentrations of engineered nanomaterials: review of modeling and analytical studies. *Environ Pollut* 2013, 181:287–300.
24. Feizi M, Jalali M, Renella G: Nanoparticles and modified clays influenced distribution of heavy metals fractions in a light-textured soil amended with sewage sludges. *J Hazard Mater* 2018, 343:208–219.
25. Zhao X, Liu W, Cai Z, Han B, Qian T, Zhao D: An overview of preparation and applications of stabilized zero-valent iron nanoparticles for soil and groundwater remediation. *Water Res* 2016, 100:245–266.
26. Mustafa G, Komatsu S: Toxicity of heavy metals and metal-containing nanoparticles on plants. *Biochim Biophys Acta* 2016, 1864:932–944.
27. Fernandes JP, Almeida CMR, Andreotti F, Barros L, Almeida T, Mucha AP: Response of microbial communities colonizing salt marsh plants rhizosphere to copper oxide nanoparticles contamination and its implications for phytoremediation processes. *Sci Total Environ* 2017, 581–582:801–810.
28. Sanz-Fernández M, Rodríguez-Serrano M, Sevilla-Perea A, Pena L, Mingorance MD, Sandalio LM, Romero-Puertas MC: Screening *Arabidopsis* mutants in genes useful for phytoremediation. *J Hazard Mater* 2017, 335:143–151.
29. Liu H, Ma C, Chen G, White JC, Wang Z, Xing B, Dhankher OP: Titanium dioxide nanoparticles alleviate tetracycline toxicity to *Arabidopsis thaliana* (L.). *ACS Sustain Chem Eng* 2017, 5: 3204–3213.
30. Servin AD, Pagano L, Castillo-Michel H, De la Torre-Roche R, Hawthorne J, Hernandez-Viecas JA, Loredo-Portales R, Majumdar S, Gardea-Torresday J, Dhankher OP, et al.: Weathering in soil increases nanoparticle CuO bioaccumulation within a terrestrial food chain. *Nanotoxicology* 2017, 11: 98–111.
31. Wang F: Decreased ZnO nanoparticle phytotoxicity to maize by arbuscular mycorrhizal fungus and organic phosphorus. *Environ Sci Pollut Res* 2018:1–12.
32. Del Real AEP, Castillo-Michel H, Kaegi R, Sinnet B, Magnin V, Findling N, Villanova J, Carríere M, Santaella C, Fernández-Martínez A, et al.: Fate of Ag-NPs in sewage sludge after application on agricultural soils. *Environ Sci Technol* 2016, 50: 1759–1768.
33. Rico CM, Majumdar S, Duarte-Gardea M, Peralta-Videa JR, Gardea-Torresdey JL: Interaction of nanoparticles with edible plants and their possible implications in the food chain. *J Agric Food Chem* 2011, 59:3485–3498.
34. Ma X, Wang C: Fullerene nanoparticles affect the fate and uptake of trichloroethylene in phytoremediation systems. *Environ Eng Sci* 2010, 27:989–992.
35. Gouda S, Kerry RG, Das G, Paramithiotis S, Shin HS, Patra JK: Revitalization of plant growth promoting rhizobacteria for sustainable development in agriculture. *Microbiol Res* 2018, 206:131–140.
36. Noori A, White JC, Newman LA: Mycorrhizal fungi influence on silver uptake and membrane protein gene expression following silver nanoparticle exposure. *J Nanoparticle Res* 2017, 19:66.
37. Judy JD, Kirby JK, McLaughlin MJ, McNear D, Bertsch PM: Symbiosis between nitrogen-fixing bacteria and *Medicago truncatula* is not significantly affected by silver and silver sulfide nanomaterials. *Environ Pollut* 2016, 214:731–736.
38. Hernández-Hernández H, González-Morales S, Benavides-Mendoza A, Ortega-Ortiz H, Cadenas-Pliego G, Juárez-Maldonado A: Effects of chitosan-PVA and Cu nanoparticles on the growth and antioxidant capacity of tomato under saline stress. *Molecules* 2018, 23:178.
39. Chen C, Tsyusko OV, McNear DH, Judy J, Lewis RW, Unrine JM: Effects of biosolids from a wastewater treatment plant receiving manufactured nanomaterials on *Medicago truncatula* and associated soil microbial communities at low nanomaterial concentrations. *Sci Total Environ* 2017, 609:799–806.

40. Sosan A, Svitunenko D, Straltsova D, Tsiurkina K, Smolich I, Lawson T, Subramaniam S, Golovko V, Anderson D, Sokolik A, *et al.*: Engineered silver nanoparticles are sensed at the plasma membrane and dramatically modify the physiology of *Arabidopsis thaliana* plants. *Plant J* 2016, **85**:245–257.
41. Hernandez-Viezcas JA, Castillo-Michel H, Peralta-Videa JR, ** Gardea-Torresdey JL: Interactions between CeO₂ nanoparticles and the desert plant mesquite: a spectroscopic approach. *ACS Sustain Chem Eng* 2016, **4**:1187–1192.
- Authors demonstrate that the desert plant *Prosopis juliflora velutina*, commonly named mesquite, is very strong candidate to be considered for removal of Ce ENMs from contaminated matrices. Interestingly, the use of X-ray absorption near edge structure (XANES) and micro X-ray fluorescence (μ -XRF) techniques, demonstrated that 81% of Ce in roots was in the same oxidation state of the CeO₂ NPs. Nevertheless, it was a 15-day hydroponic study; thus, studies evaluating the interaction of soil matrices and ENMs on mesquite plants need to be performed.
42. Rossi L, Zhang W, Schwab AP, Ma X: Uptake, accumulation and in-planta distribution of co-existing cerium oxide nanoparticles and cadmium in *Glycine max* (L.) Merr. *Environ Sci Technol* 2017, **51**:12815–12824.
- The interaction of CeO₂ and cadmium (Cd) as a co-contaminant were investigated. The study was conducted to somehow “mimic” what occurs in nature. Using soybean plants, authors demonstrated that the uptake of Ce significantly increased by the presence of the co-contaminant. Thus, the authors stated that the interactions between ENMs and co-contaminants alter plant physiology, which should be considered in the phytoremediation process and food security.
43. Deng Y, Petersen EJ, Challis KE, Rabb SA, Holbrook RD, Ranville JF, Nelson BC, Xing B: Multiple method analysis of TiO₂ nanoparticle uptake in rice (*Oryza sativa* L.) plants. *Environ Sci Technol* 2017, **51**:10615–10623.
44. Servin AD, Castillo-Michel H, Hernandez-Viezcas JA, Diaz BC, Peralta-Videa JR, Gardea-Torresdey JL: Synchrotron micro-XRF and micro-XANES confirmation of the uptake and translocation of TiO₂ nanoparticles in cucumber (*Cucumis sativus*) plants. *Environ Sci Technol* 2012, **46**:7637–7643.
45. Larue C, Castillo-Michel H, Stein RJ, Fayard B, Pouyet E, Villanova J, Magnin V, del Real A-EP, Trceras N, Legros S, *et al.*: Innovative combination of spectroscopic techniques to reveal nanoparticle fate in a crop plant. *Spectrochim Acta Part B Atom Spectrosc* 2016, **119**:17–24.
46. Tan W, Du W, Barrios AC, Armendariz R, Zuverza-Mena N, Ji Z, Chang CH, Zink JI, Hernandez-Viezcas JA, Peralta-Videa JR, *et al.*: Surface coating changes the physiological and biochemical impacts of nano-TiO₂ in basil (*Ocimum basilicum*) plants. *Environ Pollut* 2017, **222**:64–72.
47. Reddy PVL, Hernandez-Viezcas JA, Peralta-Videa JR, Gardea-Torresdey JL: Lessons learned: are engineered nanomaterials toxic to terrestrial plants? *Sci Total Environ* 2016, **568**: 470–479.
48. Raliya R, Tarafdar JC: ZnO nanoparticle biosynthesis and its effect on phosphorous-mobilizing enzyme secretion and gum contents in Clusterbean (*Cyamopsis tetragonoloba* L.). *Agric Res* 2013, **2**:48–57.
49. Dehkordi EH, Mosavi M: Effect of anatase nanoparticles (TiO₂) on parsley seed germination (*Petroselinum crispum* in vitro). *Biol Trace Elem Res* 2013, **155**:283–286.
50. Trujillo-Reyes J, Majumdar S, Botez CE, Peralta-Videa JR, Gardea-Torresdey JL: Exposure studies of core-shell Fe/Fe₃O₄ and Cu/CuO NPs to lettuce (*Lactuca sativa*) plants: are they a potential physiological and nutritional hazard? *J Hazard Mater* 2014, **267**:255–263.
51. Jasim B, Thomas R, Mathew J, Radhakrishnan EK: Plant growth and diosgenin enhancement effect of silver nanoparticles in Fenugreek (*Trigonella foenum-graecum* L.). *Saudi Pharm J* 2017, **25**:443–447.
52. Pérez-de-Luque A: Interaction of nanomaterials with plants: what do we need for real applications in agriculture? *Front Environ Sci* 2017, **5**:12.
53. Tolaymat T, Genaidy A, Abdelraheem W, Dionysiou D, Andersen C: The effects of metallic engineered nanoparticles upon plant systems: an analytic examination of scientific evidence. *Sci Total Environ* 2017, **579**:93–106.
54. López-Moreno ML, Cedeño-Mattei Y, Bailón-Ruiz SJ, Vazquez-Nuñez E, Hernandez-Viezcas JA, Perales-Pérez OJ, De la Rosa G, Peralta-Videa JR, Gardea-Torresdey JL: Environmental behavior of coated NMs: physicochemical aspects and plant interactions. *J Hazard Mater* 2018, **347**:196–217.
55. Medina-Velo IA, Barrios AC, Zuverza-Mena N, Hernandez-Viezcas JA, Chang CH, Ji Z, Zink JI, Peralta-Videa JR, Gardea-Torresdey JL: Comparison of the effects of commercial coated and uncoated ZnO nanomaterials and Zn compounds in kidney bean (*Phaseolus vulgaris*) plants. *J Hazard Mater* 2017, **332**:214–222.
- This article describes the physiological and nutrimental components of kidney bean (*Phaseolus vulgaris*) plants, exposed to coated (Z-COTE HP1®) and un-coated zinc oxide (ZnO) (Z-COTE®) nanomaterials (NMs). In the experimental design, authors also include bulk ZnO and Zinc chloride (ZnCl₂) compounds to compare and discuss their findings. Results indicate that both Z-COTE HP1® and Z-COTE® NMs, improved the physiological and nutritional parameters while the ionic Zn caused detrimental effects.
56. Demirer GS, Landry MP: Delivering genes to plants. *Chem Eng Prog* 2017, **113**:40–45.
57. Vittori Antisari L, Carbone S, Bosi S, Gatti A, Dinelli G: Engineered nanoparticles effects in soil-plant system: basil (*Ocimum basilicum* L.) study case. 2018. In press.
58. Du W, Tan W, Yin Y, Ji R, Peralta-Videa JR, Guo H, Gardea-Torresdey JL: Differential effects of copper nanoparticles/microparticles in agronomic and physiological parameters of oregano (*Origanum vulgare*). *Sci Total Environ* 2018, **618**: 306–312.
59. Xu J-B, Wang Y-L, Luo X-S, Feng Y-Z: Evaluation of zinc oxide nanoparticles on lettuce (*Lactuca sativa* L.) growth and soil bacterial community. *Chinese J Appl Ecol* 2017, **28**.
60. Zhang W, Dan Y, Shi H, Ma X: Elucidating the mechanisms for plant uptake and in-planta speciation of cerium in radish (*Raphanus sativus* L.) treated with cerium oxide nanoparticles. *J Environ Chem Eng* 2017, **5**:572–577.
- Authors studied the mechanisms by which CeO₂ NPs are taken up by radish plants. Spectroscopic and imaging analysis demonstrated that plants uptake both the NPs and released Ce ions. This article gives insights on the possible mechanisms and factors involved in the internalization of the different Ce compounds, which are relevant in the field of phytoremediation.