



RQR

*Réseau Québécois
en reproduction*

**14^{ème} Symposium annuel du
Réseau Québécois en reproduction
14-15 octobre 2021 (*en ligne*)**

**14th Annual Symposium of the
Réseau Québécois en reproduction
October 14-15 2021 (*online*)**

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**Programme du 14^e Symposium du Réseau Québécois en reproduction
Agenda of the 14th Symposium of the Réseau Québécois en reproduction**

Jeudi 14 octobre – Thursday October 14th

8h45-9h00	Connexion sur la plateforme Zoom <i>Connexion on Zoom platform</i>
9h00 – 9h15	Mot de bienvenue <i>Welcome</i>
9h15 – 10h30	<u>Présentations - Session I / Presentations – Session I</u> 5 présentations d'étudiant(e)s / 5 <i>student presentations</i>
10h30 – 10h45	Pause <i>Break</i>
10h45 – 11h45	Séminaire en production animale / <i>Animal production seminar</i> *** Johanne Cameron, Consultante indépendante spécialisée en production ovine « <i>Utilisation d'un programme lumineux intensif pour produire à l'année chez les ovins - Un succès québécois</i> »
11h45 – 13h00	Dîner <i>Lunch</i>
13h00 – 14h00	<u>Présentations - Session II / Presentations – Session II</u> 4 présentations d'étudiant(e)s / 4 <i>student presentations</i>
14h00 – 15h30	Pause-café - <u>session d'affiches I</u> <i>Coffee Break - <u>Poster Session I</u></i>
15h30 – 16h30	Conférencier invité / <i>Invited speaker</i> *** Stéphane Fabre, INRAE « <i>Genetic regulation of ovulation rate and ovarian follicular development in sheep</i> »
16h30 – 16h45	Remise du Prix MdC du RQR <i>RQR KT Award Presentation</i>

Vendredi 15 octobre – Friday October 15th

9h00 – 10h30	Session d'affiches II <i>Poster Session II</i>
10h30 – 11h30	Conférencière invitée / Invited speaker *** Deborah Bourc'his, Institut Curie « <i>The dichotomy of DNA methylation in spermatogenesis: protect or program</i> »
11h30 – 12h15	Atelier EDI / DEI workshop *** Lisa Greenhill, Association of American Veterinary Medical Colleges « <i>Be More than a DEI Supporter, Be an Ally</i> »
12h15 – 13h30	Dîner <i>Lunch</i>
13h30 – 15h00	<u>Présentations - Session III / Presentations – Session III</u> 6 présentations d'étudiant(e)s / 6 <i>student presentations</i>
15h00 – 15h15	Pause <i>Break</i>
15h15 – 16h15	Conférencière invitée / Invited speaker *** Bin Gu, Michigan State University « <i>2-Cell Based Genome Editing: From Embryonic Development to Disease Modeling</i> »
16h15 – 16h30	Remise des prix et mot de la fin <i>Awards presentation and closing</i>

Noter que le programme est à l'heure de l'Est (UTC-5)
Note that the meeting schedule is on Eastern time (UTC-5)

Session d'affiches I / Poster session I
14 octobre - October 14th
14h00 – 15h30

Quercetin mitigates H₂O₂-induced oxidative stress in bovine parthenogenetic embryos: in vitro cleavage assessment and reactive oxygen species quantification (#1), Ernesto Orozco-Lucerno, PhD, Universidad Autonoma de Ciudad Juarez (page 64).

Frailty considerations in pediatric lung transplantation (#6), John Johnson, MSc Student, University of Alberta (page 68).

Fibronectin Type III domain containing 5 (FNDC5) expression in bovine ovary and in vitro effect on bovine granulosa cell proliferation and steroidogenesis (#7), Mathilde Daudon, PhD student, Université de Montréal, (page 69)

Investigating the temporal control of mitotic exit in mammalian embryos (#9), Henry Brennan-Craddock, MSc student, Université de Montréal (page 71)

Développement d'une nouvelle méthode de caryotypage chez le porc (#10), William Poisson, MSc student, Université Laval (page 72)

The placenta for early identification of high-risk infants born at 29-36 weeks gestation (#16), Jonathan Charron, MSc student, Université de Montréal (page 75)

Single-cell look-seq to identify error prone transcriptional profiles in oocytes (#18), Karolina Kravarikova, MSc student, Université de Montréal (page 76)

The Forensic Science behind the Caribou (Rangifer) SNP Chip Validation (#19), Mallorie Trottier-Lavoie, MSc student, Université Laval (page 77).

Structure and assembly dynamics of kinetochore in oocyte meiosis-I (#20), Lin Yin, PhD student, Université de Montréal (page 78)

SFRP4 inhibe l'action des gonadotrophines dans les cellules de granulosa via un mécanisme GSK3 β /CTNNB1 dépendant (#35), *Michael Bérubé, PhD student, Université de Montréal (page 84).*

La voie de signalisation Slit/Robo est un antagoniste de la signalisation LH dans les cellules de la granulosa murines (#36), *Florine Grudet, PhD student, Université de Montréal (page 85).*

Effect of elevated NEFAs exposed during in vitro maturation on the cocultured porcine granulosa cells (#37), *Meihong Shi, PhD student, Université Laval (page 86).*

Cross-Species analysis of Wnt pathway involvement during preimplantation development and lineage specification in the human and the mouse (#38), *Katherine Vandal, PhD student, Université de Montréal (page 87).*

Implication of Fragile X-Related Proteins and neurotrophic factors in establishing transzonal projections (#42), *Mélodie Desnoyers, MSc student, Université Laval (page 91).*

Rôle du récepteur nucléaire LRH-1 dans le contrôle du métabolisme lipidique lors de la formation du corps jaune (#48), *Florence Gagnon, MSc student, Université de Sherbrook (page 94).*

Are Hippo pathway effectors potential key players of dairy cattle cystic ovarian disease pathogenesis? (#52), *Esdras Corrêa dos Santos, PhD student, Université de Montréal (page 96).*

Dynamique du remodelage de la chromatine et co-occurrence de motifs dans les cellules de la granulosa murine suite au signal ovulatoire (#53), *Fanny Morin, MSc student, Université de Montréal (page 97).*

Ablation in vivo du motif de liaison GATA dans le promoteur des gènes Star et Cyp19a1 (#56), *Julia Picard, MSc student, Université Laval (page 98).*

In vitro embryo production in Common Marmoset (#64), *Karina Gutierrez, Postdoctoral fellow, McGill University (page 103).*

The Role of Janus Kinase 3 (JAK3) in Later Stages of Follicular Development (#66), Amir Zareifard, MSc student, Université de Montréal (page 104).

3-D mitochondrial network organisation in porcine cumulus cells (#70), Amel Lounas, PhD student, Université Laval (page 105).

Peroxiredoxin 6 peroxidase and Ca²⁺-independent phospholipase A2 activities are essential for male mouse fertility (#71), Edrian Gabrielle Bumanlag, MSc student, McGill University (page 106).

ZEB1 inhibits Lhb transcription by blocking the stimulatory actions of GnRH and EGR1 (#73), Hailey Schultz, PhD student, McGill University (page 108).

The role of microRNA in the regulation of gap junction intercellular communication proteins (#74), Cameron Confuorti, MSc student, INRS (page 109).

Impact of the inhibition of the transcription factor FOXO3a during Toxoplasma gondii infection (#75), Andrés Felipe Díez Mejía, PhD student, INRS (page 110).

ATF3 regulates FSH synthesis in vitro but not in vivo (#78), Carlos Agustin Isidro Alonso, Postdoctoral fellow, McGill University (page 111).

Résumés des présentations par affiche / Poster presentations abstracts

Quercetin mitigates H₂O₂-induced oxidative stress in bovine parthenogenetic embryos: in vitro cleavage assessment and reactive oxygen species quantification (#1)

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Antioxidants can diminish oxidative stress, improving embryonic development. Here, we evaluated the effects of an oxidative stress inducer, hydrogen peroxide (H₂O₂), and the antioxidant quercetin (QUE; plant-derived flavonoid) supplementation, by assessing in vitro cleavage development and reactive oxygen species (ROS) levels in bovine parthenotes. The parthenotes were either untreated, exposed to H₂O₂ (85 µM), QUE (2 µM), or double-exposed. Development was assessed at 96 hours post-parthenogenetic activation. The rates were estimated for: Cleavage, 2-cell (2C), 4-cell (4C), early 8-cell (e8C: 5-8 cells), late 8-cell (L8C: 10-16 cells), and 5-16 cell (5-16C; e8C+L8C) stages. All cleaved parthenotes were stained with 2',7'-dichlorofluorescein diacetate to quantify ROS. The possible association between development and ROS levels was evaluated by correlation analysis. The H₂O₂-exposed parthenotes showed a significantly lower e8C-developmental rate than untreated (P<0.05). Similarly, H₂O₂-exposed parthenotes revealed less L8C-developmental rate than QUE-treated (P<0.05). Also, in H₂O₂-treated parthenotes, the 5-16C-rate was less than untreated (P=0.0557) and QUE-treated (P=0.0502). Concerning ROS, the H₂O₂-treated parthenotes exhibited higher quantities than QUE-treated 2C-parthenotes (P<0.05). Furthermore, ROS levels were higher in H₂O₂-exposed than in untreated (P<0.001), QUE-treated, and double-exposed in e8C-parthenotes (P<0.0001). Moreover, The H₂O₂-exposed parthenotes exhibited higher ROS levels, as compared with untreated (P<0.001), QUE-treated, and double-exposed (P<0.0001) at the 5-16C stage. Finally, the trends in the correlation analysis suggested inverse association between development and ROS levels. In conclusion, parthenogenetic cleavage and ROS levels appear inversely correlated. Quercetin inhibits H₂O₂-inflicted oxidative stress by mitigating ROS levels and increases in vitro cleavage in bovine parthenotes of five to 16 cells.